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ORIGINAL ARTICLE

Design, multistep synthesis and *in-vitro* antimicrobial and antioxidant screening of coumarin clubbed chalcone hybrids through molecular hybridization approach

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KEYWORDS

Coumarin; Chalcone; Coumarin clubbed chalcone; Hybrids; Antimicrobial; Antioxidant

Abstract In the present study we designed and synthesized 26 coumarin clubbed chalcone hybrids (1–13 and 14–26) in good yields (54.32–74.25%) and further tested for their antimicrobial and antioxidant activities considering the potential bioactivities of these two pharmacophores. All Spectroscopic techniques including FT-IR, 1 H NMR, 13 C NMR and mass were used to characterize the compounds. The antimicrobial and antioxidant activities of these compounds were performed by agar well diffusion method and DPPH free radical assay respectively. The compounds elicited considerable antimicrobial and potential antioxidant activities. Bioactivity data designated that the

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compounds 1 and 7 with no and 4-Cl substitution on the phenyl ring of coumarin clubbed chalcones with 3,4-dihydropyrimidine-2-one displayed antibacterial activity with minimum inhibitory concentration (MIC) value of 10 μ M and 17 μ M against *Staphylococcus aureus* and MIC value of 8 μ M and 13 μ M against *Escherichia coli* and antifungal activity with MIC value of 10 μ M and 11 μ M against Aspergillus niger respectively. On the other hand, coumarin clubbed chalcones with 3,4-dihy dropyrimidine-2(1H)-thione scaffold (14–26) exhibited potential antioxidant activity. Among them, compounds 22 containing electron releasing 2-OH substituent was the most active with 77.92% scavenging activity, followed by 14 and 26 (75.22% and 71.32%). The promising leads evolved through this investigation are important for the future development of novel and potential antioxidant compounds.

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1. Introduction

Molecular hybridization (MH) is one of the commonly employed drug design strategy by medicinal chemists for the design and development of new lead compounds. This approach will yield novel molecules that can interact with multiple sites through unique molecular interactions. MH tactic will potentiate the biological activity of the newly generated pharmacophores and reduce the side effects associated with the individual components ([Bruch et.al., 2011](#page-13-0)). In addition, MH can lead to generation of compounds with altered pharmacokinetic profiles, selectivity, dual and/or different mechanism of actions as well as the greater biological activity ([Viegas-Junior et al., 2007](#page-15-0)). Furthermore, the molecular hybrids produced are rigid scaffolds that have effective interactions with the target.

Coumarins are a type of natural products and are chemically 2H-chromen-2-one derivatives that belongs to a class of natural products known as benzopyrones [\(Venugopala et.al.,](#page-15-0) [2013, Menezes et.al., 2019, Barot et al., 2015](#page-15-0)). This scaffold is present in drugs including vitamin K antagonistic anticoagulants-warfarin, dicumarol, acenocoumarol and DNA gyrase inhibitor antibiotic-novobiocin (cathomycin/albamycin) ([Ufer, 2005, Cesar et.al., 2004, Holbrook et.al.,](#page-15-0) [2005, Gellert et.al., 1976, Sugino et.al., 1978](#page-15-0)). Chalcones are another type of natural products containing $C_6-C_3-C_6$ arrangement and are a kind of open chain flavonoids. Chalcone scaffold has attracted the scientists across the globe and many reviews have highlighted the biological and synthetic utility of these compounds ([Yazdan et.al., 2015, Gaonkar et.al.,](#page-15-0) [2017, Zhuang et al., 2017, Habib, 2018](#page-15-0)). Chalcone scaffold is found in clinically approved drugs including metochalcone

Scheme 1 Synthetic protocol for the preparation of target coumarin clubbed chalcone hybrids (1–13 and 14–26).

Fig. 1 Structures of clinically useful coumarin and chalcone derivatives.

Fig. 2 Design strategy for coumarin clubbed chalcone hybrids.

Compound Code	Chemical Structure/Formula	Appearance	$\frac{0}{0}$ Yield	Melting Point $(^{\circ}C)$	Spectral data
$U\mathbf{D}$	OН H_3C NΗ	Colorless crystals	85	$112 - 114$	MS $[m/z, \%]$: 247 (M + 1, 14.8). FT-IR (cm ⁻¹ , KBr): v 1671.24 (C=O), 1734.41 (C=O), 2925.58 (C-H), 3060.07 (Ar-C-H), 3370.82 (N-H), 3479.58 (O-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 28 (s, 3H, C-H), 2. 35 (s, 3H, C-H-C=O), 4. 91 (s, 1H, C-H), 6. 83–7.86 (m, 5H, aromatic), 9.03 (s, 1H, O–H), 9.71 (s, $1H, O-H$). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 41. 8 (C-H), 27. 3 (Me), 19. 4 (Me), 106. 2-128. 7 (Ar-C), 145. 4 (CN), 154. 2 (C-O), 150. 7 (C=O), 192. 5 (C=O).
CUD	OH H_3C NΗ H_3C Ö OH	Brick red crystals	72	$102 - 104$	MS $[m/z, \%]: 315 (M + 1, 18.1)$ FT-IR $(cm^{-1}$, KBr): v 1665.38 (C=O), 1732.21 (C=O), 2928.32 (C-H), 3038.55 (Ar-C-H), 3362 (N-H), 3456.29 (O-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 23 (s, 3H, C-H), 2. 32 (s, 3H, C-H), 5. 09 (s, 1H, C-H), 6. 82-7. 69 (m, 5H, aromatic), 9. 06 (s, 1H, N-H), 9. 88 (s, 1H, $O-H$). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 42. 3 (C-H), 19. 27. 40 (Me), 71 (Me), 91. 6–128. 2, (Ar–C), 145. 1 (CN), 150. 2, 162. 1, 192. 3 (C=O), 166.8 (C-O)
\boldsymbol{l}	O NН H_3C $C_{23}H_{18}N_2O_5$	Yellow solid	66	$91 - 93$	MS $[m/z, \%]: 403 (M + 1, 25.6).$ FT-IR $(cm^{-1}, KBr):$ v 1057.01, 1282.03, (C-O-C), 1474.89, 1601.26 (C=C), 1671.61 (C=O), 1734.27 $(C=0)$, 2925.46 $(C-H)$, 3060.24 $(Ar-C-H)$, 3370.44 $(O-H)$, 3479.20 (N-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 50 (s, 3H, C-H), 4. 95 (s, 1H, C-H), 6. 91 (s, 2H, N-H) 7. 42-7.65 $(m, 9H, Ar), 8.32$ (d, 2H, H-C=C-H), 9.11 (s, 1H, $O-H$). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 42. 5 (C–C–H), 17. 7 (CH ₃), 117. 4, -154. 8 (Ar–C), 142. 2 (C=C), 150. 2, 162, 2, 193, 3 (C=O), 166, 1(CO).
$\overline{\mathbf{c}}$	ОH Ö O NΗ Č٥ H_3C H_3CC $_{\rm H}^{\rm N}$ $C_{24}H_{20}N_2O_6$	Dark red solid	57	$107 - 109$	MS $[m/z, \%]$: 433 (M + 1, 26.4). FT-IR $(cm^{-1}, KBr): v 830.66$ (o-sub), 1024.16, 1252.28 $(C=O-C)$, 1474.34, 1611.30 $(C=C)$, 1665.06 $(C=O)$, 1732.89 (C=O), 2840.59 (C-H), 2928.48 (Ar-C-H), 3362.42 (O-H), 3436.40 (N-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 54 (s, 3H, C-H), 3.78 (s, 3H, C-H), 4.91 (s, 1H, C-H), 6.9 (s, 2H, N-H), 7. 34-7. 84 (m, 8H, Ar), 8. 37 (d, 2H, $H-C=C-H$, 9.11 (s, 1H, O-H). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 54. 3 (C-H), 42. 2 (C-C-H), 17. 9 (CH ₃), 91. 4, -159. 2 (Ar-C), 142. 6 $(C=C)$, 150. 4, 162. 6, 193. 5 $(C=O)$, 166. 5 $(C=O)$.
3	OH $\ddot{\mathrm{o}}$ NΗ H_3C $C_{24}H_{20}N_2O_5$	Yellow Solid	54	$98 - 100$	MS $[m/z, %]$: 429 (M + 1, 27.5). FT-IR (cm^{-1} , KBr): v 1057.08, 1256.20 (C-O-C), 1602.23, 1474.92 (C=C), 1674.44 (C=O), 2923.58 $(C-H)$, 3038.25 (Ar-C-H), 3269.81 (O-H), 3462.92 $(N-H)$. ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 48 (s, 3H, C-H), 4. 74 (s, 1H, C-H), 6. 12 (s, 2H, N-H), 6. 62–7. 41 (m, 9H, Ar), 7. 82 (d, 4H, C=C), 9. 05 (s, 1H, O-H). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 42. 1 (C-C-H), 17. 6 (Me), 91. 2–154. 2 (Ar–C), 141. 3, 151. 4 (C=C), 150. 1, 162. 4 193. 2 (C=O), 166. 6 (C-O).
4		Green solid	70	$135 - 137$	MS $[m/z, %_0]$: 448 (M + 1, 25.3). FT-IR $(cm^{-1}, KBr): v 756.09$ (o-sub), 1236.52 (C-O-C), 1384.47, 1531.29 (NO ₂), 1608.38 (C=C), 1672.94 (C=O), 2925.35 (C-H), 3042.09 (Ar-C-H), 3378.02 (O-H),

Table 1 Physical and spectral properties of coumarin-chalcone hybrids (1–13).

Table 1 (continued)

Table 1 (continued)

	Table 2 Physical properties and spectral data of hybrids 14–26.				
Compound Code	Chemical Structure/Formula	Appearance	$\frac{0}{0}$ Yield	Melting Point $(^{\circ}C)$	Spectral data
TUD	O OН H_3C' NН $C_{13}H_{14}N_2O_2S$	Cream colour crystals	74	$173 - 175$	MS $[m/z, \%]: 263 (M + 1, 12).$ FT-IR (KBr, cm ⁻¹): v 1275.21 (C-O), 1472.97, 1602.84 (C=C), 1671.10 (C=O), 2925.07 (C-H), 3021.42 $(Ar-C-H)$, 3420.82 (O-H), 3248.73 (N-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 1. 35 (s, 3H, C-H), 2. 03 (s, 2H, N-H, Ar), 2. 82 (s, 3H, $C-H-C=O$, 4. 96 (s, 1H, $C-H$), 5. 21 (s, 1H, $O-H$), 6. $56-6.95$ (m, 4H, Ar). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 42.5 (C–H, Ar), 27.2 (Me), 17.8 (Me), 104.6-128.6 (Ar-C), 154. 6 $(C-O-H)$, 158. 1 $(C-N)$, 174. 5 $(C=S)$, 196. 3 $(C=O)$.
CTUD	OН Ω NΗ H_3C $C_{23}H_{18}N_2O_4S$	Red crystals	81	$111 - 113$	MS $[m/z, %0]$: 331 (M + 1, 9). FT-IR (cm ⁻¹ , KBr): v 1475.26 (C=C), 1612.04 (C-O), 1665.93 (C=O), 2930.11 (C-H), 3016.77 (C-H Ar), 3248.29 (N-H), 3418.82 (O-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 1. 26 (s, 3H, C-H), 2. 08 (s, 2H, N-H, Ar), 2. 62 (s, 3H, C-H-C=O), 4. 74 (s, 1H, C-H), 6. 76-7. 10 (m, 4H, Ar), 9. 35 (s, 1H, $O-H$). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 43.7 (C-H, Ar), 27.6 (Me), 15.4 (Me), 91.6–148.2 (Ar–C), 158. 4 (CN), 166. 2 (C-O), 174. 1 (C=S), 160. 5, 196. 7 (C=O).
14	OН O NΗ `S H_3C $C_{23}H_{18}N_2O_4S$	Yellow solid	69	$106 - 108$	MS $[m/z, \%]$: 419 (M + 1, 11). FT-IR (cm ⁻¹ , KBr): v 1039.60, 1232.87 (C-O), 1490.49, 1608.91 (C=C), 1671.08 (C=O), 2850.79 (C-H), 2919.91 (Ar-C-H), 3347.96 (N-H), 3472.96 (O-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 42 (s, 3H, C-H), 4.85 (s, 1H, C-H), 6.35 (s, 2H, N-H), 6.37–7.33 (m, 9H, Ar), 7.89 (d, 2H, C=C), 9.12 (s, 1H, O-H). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 43.7 (C–C–H), 17.2 (Me), 91.5–148.5 (Ar–C), 166.1 (C–O–H), 167.8 $(C-N)$, 174.4 $(C=$ S), 160.6, 193.8 $(C=$ O).
15	OH O NН H_3CC H_3C $C_{24}H_{20}N_2O_5S$	Dark red solid	61	$146 - 148$	MS $[m/z, \%]$: 449 (M + 1, 10). FT-IR $(cm^{-1}, KBr): v 759.79$ (o-sub), 1050.02, 1287.92 (C-O), 1425.17 (C=C), 3424.04 (O-H), 1602.27 (C=C), 1682.31 (C=O), 2948.84 (C-H), 3055.15 (Ar-C-H), 3318.93 (N-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 38 (s, 3H, C-H), 3. 14 (s, 3H, C-H), 4. 66 (s, 1H, C-H), 6. 11 (2H, N-H), 6. 69-7. 21 (m, 8H, Ar), 7. 61 (d, 2H, C=C), 9.58 $(s, 1H, 0-H).$ ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 55.4 (Me), 42.9 $(C-C-H)$, 15.8 (Me), 91.2-148.5 (Ar-C), 152. 5 (C=C), 158. 2 (C-OCH ₃), 166. 8 (C-O-H), 167. 4 (C-N), 174. 1 $(C=$ S), 160.7, 193.2 $(C=$ O).
16	OH NΗ H_3C $C_{25}H_{20}N_{2}O_{4}S$	Yellow solid	58	$142 - 144$	MS $[m/z, \%]$: 445 (M + 1, 8). FT-IR $(cm^{-1}, KBr): v 753.87 (Ar), 1411.10 (C=C),$ 1688.83 (C=O), 2893.03 (C-H), 3013.90 (Ar-C-H), 3218.17 (N-H), 3415.38 (O-H). ¹ H NMR (400 MHz, DMSO d_6 , ppm): 2. 56 (s, 3H, C-H), 4.85 (s, 1H, C-H), 6.15 (s, 2H, N-H), 6.91-7.35 (m, 9H, Ar), 7. 55 (d, 2H, C=C), 9. 47 (s, 1H, O-H). ¹³ C NMR (100 MHz, DMSO d_6 , ppm): 43.7 (C–C–H), 17.3 (Me), $91.8-135.4$ (Ar–C), 131. 5, 151. 5 (C=C), 166. 2 (C-O), 167. 9 (C-N), 174. 7 (C=S), 160.3, 193.4 $(C=0)$.
17		Green solid	66	$152 - 154$	MS [m/z , %]: 464 (M + 1, 8). FT-IR $(cm^{-1}, KBr): v 783.91 (p-sub), 1045.88, 1282.05)$ (C-O), 1445.24 (C=C), 1382.15, 1535.04 (NO ₂), 1684.37 (C=O), 2938.22 (C-H), 3019.29 (Ar-C-H), 3284.49 (continued on next page)

Table 2 (continued)

Table 3 Antimicrobial activity of coumarin clubbed chalcones against selected bacterial and fungal strains.

Sa: Staphylococcus aureus; Ec: Escherichia coli; An: Aspergillus niger; $R =$ Resistance.

and sofalcone [\(Fig. 1\)](#page-2-0) that are used as choleretic and antiulcer agents [\(Aksoz and Ertan, 2013, Sahu et.al., 2012, Shigeru et.](#page-13-0) [al.,1991\)](#page-13-0). Additionally, clinical trials data indicated that hesperidin trimethylchalcone and hesperidin methylchalcone screened for branch or trunk varicosis and chronic venous lymphatic insufficiency respectively were biologically effective due to their ability to reach required concentrations in plasma, reducing the symptoms and good acceptance by the physiological system ([Liu et.al., 2016, Beltramino et.al., 2000, Guan et.](#page-14-0) [al., 2014](#page-14-0)).

Many scientists synthesized and screened various coumarin hybrids as well as the chalcone hybrids with potential bioactivities. Molecular hybrids of coumarins and chalcones possess excellent antibacterial [\(Feng et.al., 2020, Osman et.al.,2018,](#page-13-0) Sanad and Mekky, 2020, Kraljević et.al., 2016, Sahoo et al., [2021, Lipeeva et.al.,2019, Vazquez-Rodriguez et.al., 2015,](#page-13-0) [Sashidhara et.al., 2015, Shaik et.al., 2017, Wang et al., 2019b,](#page-13-0) [Sribalan et al., 2016\)](#page-13-0), antifungal ([Zhang et al., 2021, Yang and](#page-15-0) [Liang, 2021, Patel et al., 2017, Sharma and Katiyar, 2019,](#page-15-0) [Prusty and Kumar, 2020; Mellado et al., 2020, Singh et al.,](#page-15-0) [2018, Shaik et al., 2020b, Kant et al., 2016, Lagu et al., 2020,](#page-15-0) [Akkulu Naidu and Rajendra Prasad, 2018\)](#page-15-0), antioxidant ([Matos et al., 2015, Nagamallu et al., 2016, Karina et al.,](#page-14-0) [2018, Shi et al., 2020, S](#page-14-0)[enocak et al., 2018, Aneja et al., 2018,](#page-14-0) Jakovljević [et al., 2018, Xue et al., 2018, Latif et al., 2020, Arif](#page-14-0) [et al., 2020, Kostopoulou et al., 2020](#page-14-0)) and anticancer activities ([Zhang and Xu, 2019, Kurt et al., 2020, Kamath et al., 2015,](#page-15-0) [Xu et al., 2019, Song et al., 2020, Goud et al., 2019, Gao et al.,](#page-15-0) [2020, Park et al., 2018, Wang et al., 2019a, Mirzaei et al.,](#page-15-0) [2020, Djemoui et al., 2020\)](#page-15-0) etc. Coumarins and chalcones comprise antioxidant activity due to their ability to influence the formation and scavenging of reactive oxygen species (ROS) (Fylaktakidou et al., 2004, Sökmen and Khan, 2016). Similarly, a variety of mechanisms are responsible for the antibacterial and antifungal activities of coumarins and chalcones [\(Qin et al.,](#page-14-0) [2020, Srikrishna et al., 2018, Hu et al., 2018, Dan and Dai,](#page-14-0) [2020, Farhadi et al., 2018, Xu et al., 2019, Wei et al., 2016\)](#page-14-0). Coumarin containing hybrids like clorobiocin, coumermycin A1 as well as novobiocin are already employed clinically to treat different bacterial infections. Hence, it is possible to link coumarin moiety with another antimicrobial pharmacophore like chalcone via the molecular hybridization strategy to yield novel drugs against infectious diseases. Coumarins and chalcones can be conveniently synthesized in the laboratory through Pechmann and Claisen-Schmidt condensation reactions ([Gaudino](#page-13-0) [et al., 2016, Shaik et al., 2019, Chavan et al., 2017, Vijaya](#page-13-0) [Bhargavi et.al., 2017](#page-13-0)).

Bearing in mind the synthetic feasibility and promising bioactivities of hybrids derived from coumarins and chalcones, in the present investigation we prepared ([Scheme 1\)](#page-1-0) and screened the antimicrobial and antioxidant properties of novel coumarin-clubbed chalcone derivatives ([Fig. 2\)](#page-2-0).

2. Materials and methods

2.1. General

The chemicals and reagents employed for the synthesis were purchased from local suppliers and the melting points of the prepared compounds were determined using Dalal melting point apparatus in open capillary tubes. FT-IR spectra were scanned by KBr-Pellet method. ${}^{1}H$ NMR and ${}^{13}C$ NMR spectra and mass spectra were recorder at Laila Impex, Vijayawada, Andhra Pradesh, India by using a Bruker-NMR Spectrometer 400 MHz and Shimadzu GCMS QP 5000 mass spectrometers respectively. Other chemicals were of commercial grade and used without any purification further. The reactions were monitored by thin-layer chromatography (TLC) on silica gel F_{254} aluminium TLC plates (Merck, Germany) using ethyl acetate (EtOAc)-hexane mixture as mobile phase. The spots on the TLC plate were visualized using iodine vapours and UV lamp.

2.2. Experimental

2.2.1. Synthesis of 5-Acetyl-4-(2-hydroxyphenyl)-6-methyl-3,4 dihydropyrimidin- $2(1H)$ -one (UD)

2-Hydroxybenzaldehyde (1 mmol), 1.1 mmol of pentane-2,4 dione and 1.3 mmol of urea were refluxed and stirred in 100 mL round bottom flask (RBF) at 80 $\,^0C$ in the presence of 0.5 mmol of citric acid as catalyst. After the end of the reaction (that is monitored through TLC 30% EtOAc: Hexane mixture), the reaction mixture was charged with 100 mL of cold water and stirred for another 10 min. This led to the formation of a precipitate which was further filtered and thoroughly washed with distilled water and dried in vacuum to get the crude solid that was purified further by recrystallization using ethanol (Yield, 85%) [[Hajelsiddig and Saeed, 2015\]](#page-14-0).

2.2.2. Synthesis 5-Acetyl-4-(4-hydroxy-2-oxo-2H-chromen-8 y l)-6-methyl-3,4-dihydropyrimidin-2(1H)-one (CUD)

To a mixture of 5-Acetyl-4-(2-hydroxyphenyl)-6-methyl-3,4-d ihydropyrimidin-2(1H)-one (0.1 mol) and malonic acid (0.1 mol), 40 mL of phosphorus oxychloride and 30 g of anhydrous zinc chloride (preheated to $60-70$ °C) were added. The above mixture was heated for 20 h at 70 \degree C and after the completion of the reaction that was monitored by TLC (40% EtOAc-Hexane mixture), 50 mL of ice-cold water was added to the reaction mixture to form a precipitate. The formed precipitate was washed thoroughly with cold water and then filtered. The solid thus obtained was treated with 10% sodium carbonate solution and filtered and to the filtrate diluted hydrochloric acid was added to obtain crude product. The crude product was filtered, washed with water, dried and recrystallized using glacial acetic acid (Yield, 74%) [\[Potdar](#page-14-0) [et al., 2005\]](#page-14-0).

2.2.3. General procedure for synthesis coumarin clubbed chalone derivatives (1–13)

5-Acetyl-4-(4-hydroxy-2-oxo-2H-chromen-8-yl)-6-methyl-3,4 dihydropyrimidin-2(1H)-one (0.002 mol) and substituted aromatic aldehyde (0.002 mol) were dissolved in 6 mL of ethanol. Then, 3 mL of 40% KOH was added drop wise to the above solution and the reaction mixture was stirred for 30 h. The completion of the reaction was monitored by TLC (30% EtOAc-Hexane for most of the compounds. However, for compounds 5, 9 and 13, 40% EtOAc-Hexane was used whereas for compound 6, 20% EtOAc-Hexane was employed as mobile phase). The reaction mixture was added with crushed ice and the contents were neutralized with dil. HCl to obtain a precipitate. The crude solid formed was recrystallized using ethanol (95%) [[Konidala et al., 2021; Lokesh](#page-14-0) [et al., 2017; Shaik et al., 2015](#page-14-0)]. Physical properties and spectral data of hybrids 1–13 are displayed in [Table 1](#page-3-0).

2.2.4. General procedure for synthesis of 1-(4-(2- Hydroxyphenyl)-6-methyl-2-thioxo-1,2,3,4 tetrahydropyrimidin-5-yl)ethanone (TUD)

TUD was synthesized and purified employing the same protocol as described under the general procedure for the synthesis of UD except that thiourea is replaced with urea.

2.2.5. General procedure for synthesis 8-(5-acetyl-6-methyl-2 thioxo-1,2,3,4-tetrahydropyrimidin-4-yl)-4-hydroxy-2Hchromen-2-one (CTUD)

CTUD was synthesized and purified employing the same protocol as described under the general procedure for the synthesis of CUD except that 1-(4-(2-Hydroxyphenyl)-6-methyl-2 thioxo-1,2,3,4-tetrahydropyrimidin-5-yl)ethanone was used replacing 5-Acetyl-4-(2-hydroxyphenyl)-6-methyl-3,4-dihydro pyrimidin-2(1H)-one.

2.2.6. General procedure for synthesis coumarin clubbed chalone derivatives (14-26)

The same protocol employed for the synthesis of compounds 1–13 is utilized for the synthesis of the target compounds 14– 26. However, in this case, CTUD was used as a starting material in the place of CUD and the reactions were monitored by TLC employing 50% EtOAc-Hexane as mobile phase [[Konidala et.al., 2020](#page-14-0)]. Physical properties and spectral data of hybrids 14–26 were given in [Table 2.](#page-6-0)

2.3. In vitro Anti-microbial activity:

The in vitro antimicrobial testing was performed by Agar well diffusion method [\[Bauer et.al., 1966, Lagu et.al., 2019](#page-13-0)] against Staphylococcus aureus (NCIM 2122), Escherichia coli (NCIM 2137) and Aspergillus niger (NCIM 652). The zone of inhibition of target compounds was measured and compared with standard drugs including ciprofloxacin and fluconazole. The microbial strains were sub-cultured in presterilized nutrient agar and potato dextrose agar media for antibacterial and antifungal activity respectively. Then the pre-inoculated medium was aseptically transferred into sterilized Petri plates of 4-inch diameter. After the solidification of the medium, sterilized cork borer was used make a cup of 6 mm and labelled. The solutions of the standard compound in concentration of 10 lg/ml was prepared in DMSO where as the solutions of the test compounds were obtained by dissolving in DMSO and diluting to a dose level of 25, 50, 75 and 100 μ g/ml. These concentrations were later added to the bore of Petri plates under sterilized conditions, and the plates were incubated for 48 h. DMSO was used as a negative control and found no

effect. The Zone of inhibition was determined and MIC values were calculated for each test compound along with standard. The results of antibacterial activity are tabulated in [Table 3.](#page-9-0)

2.4. In vitro antioxidant activity

The percentage of antioxidant activity (AA^{ϕ}) of each substance was determined by DPPH free radical assay method. DPPH radical scavenging activity [\[Shaik et al., 2020a\]](#page-15-0) of the samples was assessed by reacting with stable DPPH radical in an ethanol solution. The reaction mixture comprises 0.5 mL of sample, 3 mL of absolute ethanol and 0.3 mL of 0.5 mM DPPH radical solution in ethanol. The change in the color from deep violet to light yellow due to the reaction of DPPH with the target compound was read at 517 nm after 30 min of reaction using a UV–VIS spectrophotometer. The mixture of ethanol (3.3 mL) and sample (0.5 mL) serve as blank whereas the control solution was prepared by mixing ethanol (3.5 mL) and DPPH radical solution (0.3 mL). The scavenging activity percentage (AA%) was determined according to previously reported method [\[Mensor et al., 2021](#page-14-0)].

$$
-\left[\frac{(Absorbance of the sample - Absorbance of Control)}{Absorbance of Control}\right]
$$

× 100

3. Results and discussion

3.1. Chemistry

The acid catalyzed Biginelli reaction of salicylaldehyde with pentane-2,4-dione and urea/thiourea afforded the dihydropyrimidine derivatives, UD and TUD in 85.41% and 74.38% yields respectively. The characteristic absorption bands for the secondary amino group $N-H$ at wave numbers 3370 and 3248 cm⁻¹ and singlet peaks in the 1 H NMR for methine (CH) proton at the chemical shifts 4.91 and 4.96 ppm confirmed the formation of UD and TUD. Further Pechmann condensation of UD and TUD with malonic acid, phosphorous oxychloride and zinc chloride formed CUD (72.45%) and CTUD (81.66%) in good yields. The mass spectrum of these two compounds showed $M + 1$ peak at m/z values 315 and 331 corresponding to their molecular weights. Additionally, the FT-IR, ${}^{1}H$ NMR, and ${}^{13}C$ NMR spectrum of all

the compounds and the mass spectrum of UD and TUD showed characteristic bands and peaks and established the structures of UD, TUD, CUD and CTUD. The target coumarin clubbed chalcone hybrids (1–13 and 14–26) were then synthesized by the base catalyzed Claisen-Schmidt condensation of CUD and CTUD with a variety of aromatic aldehydes in the yields ranging from 54.32 to 74.25%. These compounds showed the characteristic absorption bands in their FT-IR spectrums corresponding to the $-CH=CH-$ and $-C=O$ functional groups of the ketovinyl fragment of chalcones. Additionally, these hybrids also displayed characteristic peaks in their ${}^{1}H$ and ${}^{13}C$ nuclear magnetic resonance spectra's and the molecular ion peaks at their respective m/z values as $M + 1$ and $M + 2$ peaks aided in dereplicating the structure of the proposed target compounds.

3.2. Antimicrobial activity

The antimicrobial screening data of the two series of compounds (1–13 series and 14–26 series) is depicted in [Table 3](#page-9-0). Ciprofloxacin and Fluconazole were used as reference standard for the assay. In addition, between these two series compounds belonging to 1–13 series contain oxygen in the pyrimidine ring whereas the 14–26 series compounds possess sulfur atom. Among the two series, compounds belonging to 1–13 series possess considerable antimicrobial activity and the organisms showed resistance for majority of compounds in 14–26 compounds series. Overall, the MIC values of the compounds are in the range of $8-49 \mu M$. In 1–13 series, it can be seen that compound 1 (MICs $8-10$) and 7 (10– $17 \mu M$) containing unsubstituted phenyl and 4-chlorophenyl ring showed better antimicrobial activity over other compounds (Fig. 3). However, despite of changing the electronic properties of the phenyl ring, no improvement was observed in the MIC values against reference standard.

3.3. Antioxidant activity

The antioxidant activity [\(Table 2\)](#page-6-0) indicated by the $\%$ scavenging activity of synthesized compounds (1–13 and 14–26) was determined by the DPPH assay method, data is depicted in [Table 4](#page-12-0). Ascorbic acid (AA) and Butylated hydroxytoluene (BHT) were used as reference standard. The compounds and the standards were evaluated at a test concentration of 100 mg/ml. The compounds belonging to 14–26 series showed better antioxidant activity the compounds belonging to 1–13

Fig. 3 Coumarin clubbed chalcone hybrids with considerable antimicrobial activity.

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Fig. 4 Potential coumarin clubbed chalcone hybrids with antioxidant activity.

4. Conclusions

series. In 1–13 series, the best antioxidant activity was observed for compound 5 at 66%. The compounds 5, 14, 22 and 26 showed significant antioxidant activity with % scavenging activity in the range of 66.21–77.92 %. Among these, 14, 22 and 26 containing unsubstituted phenyl ring and phenyl ring substituted with electron releasing 2-OH substitution and 3,4,5-trimethoxy substituents in chalcones with 3,4-dihydropyr imidine-2(1H)-thione moiety showed excellent antioxidant activity with 75.22%, 77.92% and 71.32% scavenging activity compared to BHT (Fig. 4). However, their activity was less but close to AA.

Two series (1–13 series and 14–26 series) having a total of 26 new coumarin clubbed chalcone hybrids were synthesized, characterized and evaluated for their antimicrobial and antioxidant activities. The antimicrobial screening of all the synthesized compounds was done by in-vitro well diffusion method against Gram $+$ ve, Gram -ve, and fungal strains, the results concluded that the compounds 1 and 7 bearing phenyl and 4-Cl phenyl substitution showed potent antibacterial activity

with MIC value of 10 μ M and 17 μ M against Gram + ve (*Sta* $phylococcus$ aureus) and MIC value of 8 μ M and 13 μ M against Gram –ve (Escherichia coli) and antifungal activity having MIC value of 10 μ M and 11 μ M against *Aspergillus* niger respectively. The antioxidant activity screening was carried out by in-vitro DPPH assay; results indicated that the compound 22 showed the most potent antioxidant activity with $\%$ scavenging activity as 77.92%. The antioxidant activity of the designed compounds is promising than the antimicrobial activity. Hence, the potential antioxidant compounds emerged from the structure activity relationship studies would be utilized for future research for the design of newer antioxidant analogs of coumarin clubbed chalcone hybrids.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary material

Supplementary data to this article can be found online at [https://doi.org/10.1016/j.arabjc.2021.103154.](https://doi.org/10.1016/j.arabjc.2021.103154)

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